



Title: *Collection of Bronchoalveolar (BAL) Cells*

No: RTLP-GL-CP1

Location:
Old CCRC Tripp Lab

Approval Date:
10 September 2004

Supersedes Date:

Materials:

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| •Lab coat | •Phosphate Buffered Saline (PBS) | •Benchcoat | •50 mL tubes |
| •Gloves | •Dissecting scissors & forceps (2) | •Ice bucket w/ice | •BSA |
| •Mask | | •15mL tubes | •1 mL syringe (2) |
| •Plastic cannula | | •Avertin | •25 gauge needle |

Procedure:

1. Hold mouse in dominant hand by the scruff of neck and loop pinky-finger around base of the tail. Hold firmly and turn mouse so ventral side is facing upward.
2. Inject 300 cc avertin solution (RTLP-GL-BSS-30) into peritoneal cavity of mouse and place mouse in a clean cage and wait for it to lie calmly.
3. Once mouse is sedated, place onto benchcoat paper and snip the right caudal artery. Lay mouse on it's ventral side to allow blood to drain. NOTE: If collecting serum, use transfer pipette to harvest and place into sterile 1.5 mL microfuge tubes.
4. Once blood has drained sufficiently, lay mouse spleen-side up and snip the fur and make a v-shaped opening, being careful not to pierce peritoneal cavity. Pinch the fur with fingers and pull fur off and over the head of the mouse.
5. Clean area around the trachea with forceps, then snip the trachea near hyoid bone (nearest to the cervical region).
6. Using a fresh syringe, draw up 1mL of PBS+1% BSA buffer and insert tip of plastic cannula into the tracheal opening.
7. Triturate three times and place collected cells into 15mL tube (on ice).
8. Repeat steps 6 and 7 twice more for a total of three times.
9. Wrap up mice, place into plastic bag, and put in mouse storage freezer for animal resources to pick up.

| Author | Management Approval/Date | Effective Date |
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